

# Introduction to Bioinformatics

## 6. DNA Restriction Analysis and Primer Design

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Laboratory

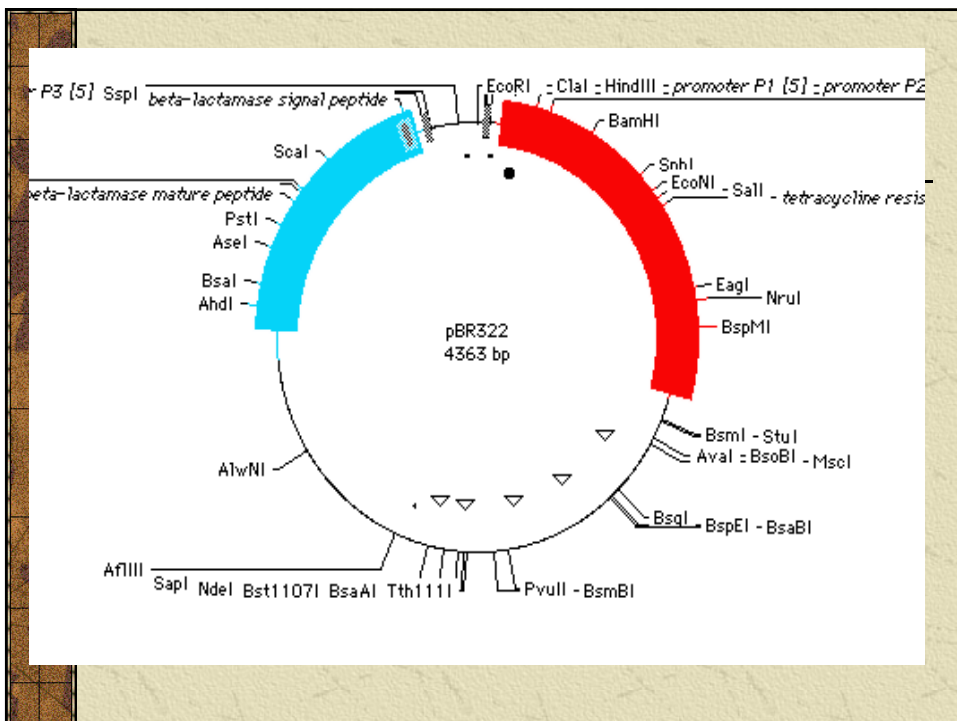
Beltsville, MD 20708

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### What we will cover today

- ☐ Restriction site analysis
  - ☐ Cloning
  - ☐ Southern blot analysis
- ☐ PCR primer design
  - ☐ Cloning, amplification gene expression

## ✳ Southern blot interpretation



## Restriction Mapping

- ✠ Making restriction maps was my first use of “Molecular Biology” software
- ✠ Making restriction maps is a routine lab activity that is necessary for any type of cloning project.
- ✠ High quality maps are important for publications and exchange of information between researchers or between labs.

## Mapping Software

- ✠ Programs vary greatly in sophistication and ease of use
  - ◆ Simple drawing programs (vector graphics)
  - ◆ The venerable DNA Strider
  - ◆ GCG (not a strong point of the package)
  - ◆ Comprehensive Mac/PC MolBio programs
  - ◆ Dedicated plasmid drawing programs
  - ◆ Can it be done on the Web?
- ✠ Making high quality graphical restriction maps is one area where Mac/PC programs are much better than GCG or the Web



# Bitools@UMASS Medical School

✧ <http://biotools.umassmed.edu/tacg/WWWtacg.php>

✧ Enter your DNA sequence

✧ Select limits


BioTools: WWWtacg Ver 3.0 - Microsoft Internet Explorer

File Edit View Favorites Tools Help

Back Forward Stop Home Search Favorites Media Mail Print

Address <http://biotools.umassmed.edu/tacg/WWWtacg.php> Go Links

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 **BioTools @ UMass Medical School** Research Computing

Biotools -> BioApps

Tue, Aug 03 Home | News | Resources | Contact 11:54:11

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### TACG Restriction Mapping

This form allows you to supply both DNA sequence and (optionally) your own file of Restriction Enzymes or other IUPAC patterns **in GCG format** (or slightly modified for more functionality) for Restriction Enzyme Mapping and Analysis, using **Harry Mangalam's tacg3.5** program as the analysis engine.

<----- You should be able to see the ends of this line in your window ----->

Most of the options are linked to more help. If needed; you can also look at the [local HTML-ized tacg man page](#) and [local tacg documentation](#) for more information.

PLEASE [mail me](#) if you notice something odd or objectionable in presentation, execution, or especially (!) accuracy of results.

**Sequence Entry**

**Any ASCII format** is acceptable; tacg3 accepts and processes **all IUPAC degeneracies**.  
With the assistance of Jim Knight's SEQIO pkg (now incorporated into tacg), WWWtacg3 will also accept **MOST NUCLEIC ACID FILE FORMATS** without further modification or editing.

--EITHER--Paste in the RAW sequence (>4 bp) in the window below...

Unlabeled  
sequence

[Optional Output Label for pasted-in  
sequence]

**BioTools: WWWtacg Ver 3.0 - Microsoft Internet Explorer**

File Edit View Favorites Tools Help

Back Forward Stop Home Search Favorites Media

Address http://biotools.unmassmed.edu/tacg/WWWtacg.php Go Links

--EITHER--Paste in the RAW sequence (>4 bp) in the window below...

[Unlabeled] [Optional Output Label for pasted-in sequence]

```

2881 acgtgaattt tgcgaaat gaatgatgc gatggatgc cgaacacatg caaatttga
2941 ccactattt atttacttg attttctcg taggtattt gcttctata acgtttagt
3001 aatgaagctt gtttctctt atctttgtc tgaagaata ataaggaat ggaacagaac
3061 tccattaacc ttgattgga gttacattgt aaaggaacgg aagtaaacag aatttattt
3121 aattttaac ttgcctaact gttctttta taaaaaaaa aaaaaaa

```

--OR-- use the file browser below

NB: if there's sequence in the window above, the file will be ignored.  
 [Mac/Win32 versions of Netscape up to V4.7 (but not linux/Unix) will not accept more than ~30K chars in the paste-in buffer. If you need to load large files, save as text and load them via the file browser below]

Use the file browser below to upload a **DNA sequence file (<500,000 characters) to send:**  
 (Output will be labeled with the **file name**.)

Browse...

☐ Force sequence file to be Read as 'raw'. (only numbers, spaces, and sequence)

**Sequence Selection and Numbering Options**

The sequence should be analyzed from bases  to   
 (use "END" to indicate end; integers for other endpoints.)

The (sub)sequence should be analyzed as: ☒ Linear ☐ Circular

Sequence numbering should start from  (Only for Linear Map; Sites are numbered starting from 1.)

**BioTools: WWWtacg Ver 3.0 - Microsoft Internet Explorer**

File Edit View Favorites Tools Help

Back Forward Stop Home Search Favorites Media

Address http://biotools.unmassmed.edu/tacg/WWWtacg.php Go Links

Reset to Defaults Submit Sequence to WWWtacg

**Restriction Enzyme Selection**

Select the **Restriction Enzymes** either **By Characteristic** or **Explicitly**  
 Filtering options below apply to entire REBASE files (yours or mine), but NOT to explicitly picked enzymes

☐ Search for possible **SILENT Sites** (supercedes all other selections and forces Linear Map Mode).

By Characteristic	Explicitly																																						
This side is the default.	This side is selected if ANY names are selected																																						
<b>Overhang</b> <input type="text" value="Don't Care"/> <b>Number of cuts:</b> Minimum: <input type="text" value="1"/> Maximum: <input type="text" value="1"/> <b>Magnitude of Recognition</b> <b>Sequence:</b> <input type="text" value="6"/> base pairs or greater. <b>Cost:</b> <input type="text" value="No Limit"/> Selections filtered from from the Latest REBASE file <input type="text" value=""/> <b>--OR--</b> from YOUR uploaded <b>GCG-formatted REBASE file</b> , using the file browser below. <i>Uploads supercede the menu selection.</i>	<input type="checkbox"/> Simulate multiple digestion as well as singles. <table border="1"> <tbody> <tr><td>AatII</td><td>BmgI</td></tr> <tr><td>AccI</td><td>BpII</td></tr> <tr><td>AccII</td><td>Bpml</td></tr> <tr><td>Acil</td><td>Bpu10I</td></tr> <tr><td>AcII</td><td>Bpu1102I</td></tr> <tr><td>AdII</td><td>BsaI</td></tr> <tr><td>AdIII</td><td>BsaAI</td></tr> <tr><td>BsrGI</td><td>EcoRV</td></tr> <tr><td>BssHII</td><td>FauI</td></tr> <tr><td>BssSI</td><td>Fnu4HI</td></tr> <tr><td>Bst4CI</td><td>FokI</td></tr> <tr><td>BstAPI</td><td>FseI</td></tr> <tr><td>BstDSI</td><td>FspI</td></tr> <tr><td>BstEII</td><td>GdII</td></tr> <tr><td>NdeI</td><td>ScrFI</td></tr> <tr><td>NgoAIV</td><td>SexAI</td></tr> <tr><td>NheI</td><td>SfaNI</td></tr> <tr><td>NlaII</td><td>SfiI</td></tr> <tr><td>NlaIV</td><td>SfiI</td></tr> </tbody> </table>	AatII	BmgI	AccI	BpII	AccII	Bpml	Acil	Bpu10I	AcII	Bpu1102I	AdII	BsaI	AdIII	BsaAI	BsrGI	EcoRV	BssHII	FauI	BssSI	Fnu4HI	Bst4CI	FokI	BstAPI	FseI	BstDSI	FspI	BstEII	GdII	NdeI	ScrFI	NgoAIV	SexAI	NheI	SfaNI	NlaII	SfiI	NlaIV	SfiI
AatII	BmgI																																						
AccI	BpII																																						
AccII	Bpml																																						
Acil	Bpu10I																																						
AcII	Bpu1102I																																						
AdII	BsaI																																						
AdIII	BsaAI																																						
BsrGI	EcoRV																																						
BssHII	FauI																																						
BssSI	Fnu4HI																																						
Bst4CI	FokI																																						
BstAPI	FseI																																						
BstDSI	FspI																																						
BstEII	GdII																																						
NdeI	ScrFI																																						
NgoAIV	SexAI																																						
NheI	SfaNI																																						
NlaII	SfiI																																						
NlaIV	SfiI																																						

BioTools: WWWtacc Ver 3.0 - Microsoft Internet Explorer

File Edit View Favorites Tools Help

Back Forward Stop Search Favorites Media Print Mail Print Mail Print

Address http://biotools.umassmed.edu/tacc/WWWtacc.php Go Links

**Output Format**

The output should be 60 chars wide, in Regular Small tiny font.

**Suggestions**  
NB: Choosing the **VERY** wide option makes **most of the output** stream out in a single line.

☒ **Sort Output by # Hits, Names** (DNA Strider-style)

**For debugging strange results:**

☐ Show me the tacc **command line** that generated these results.

☐ Show me **ERRORs from tacc**.

**Analyses**

☒ **Summary Table** (Absent Sites, # of sites for each Pattern)

☐ **GCG-like Ladder Map** (req >100 b to be useful)

☐ **Pseudo Gel Map** LOW cutoff at 100 bp; HIGH cutoff at 1,000 bp.

☐ **Table of Hit Sites**

☐ **Table of Fragments** Listed by: Size

☐ **Stream out Open Reading Frames** of greater than 100 Amino Acids in frames ☒ 1 ☐ 2 ☐ 3 ☐ 4 ☐ 5 ☐ 6

☐ Print the EXTENDED ORF info.

☐ Print the ORF sequence UNWRAPPED (1 line output).

☒ **Entire Linear Map Long**, but required for Co-Translation below.

☒ Only print top strand ☐ No tic marks ☐ Print Map even if No Hits

☒ **Linear Co-Translation:** in 1 frame(s), with a 1 letter code, using Standard Codon Usage.

Reset to Defaults Submit Sequence to WWWtacc

## Results of query

### ✳ Enzymes that DO NOT MAP

◆ table

### ✳ Enzymes that cut and number of cuts

◆ table

### ✳ Linear map of sequence with cut sites and amino acid translation



Restriction Enzyme Site Mapper version 3 - Microsoft Internet Explorer

http://www.restrictionmapper.org/

## Welcome to RestrictionMapper - Now with Virtual Digest

Maps sites for restriction enzymes (restriction endonucleases) in DNA sequences.

[NEW! Automate RestrictionMapper](#)

[HELP](#)  
[FAQ](#)  
[WHAT'S NEW?](#)  
[CODE](#)  
[CONTACT](#)  
[Other Free Molecular Biology Resources](#)  
[Dilution Calculator](#)

**Conformation**

Circular ☐  
Linear ☒

**Sort By**

1. frequency  
2. overhang  
3. name

Maximum Cut Number

Minimum Site Length

**Include**

Select Individual Enzymes

All Enzymes  
AarI  
AasI  
AatI  
AatII  
AccI  
AccII

All Commercial ☒  
NEB only ☐

5' overhang ☒  
3' overhang ☒  
blunt ☒

Prototypes Only ☒  
All Isoschizomers ☐

**Sequence Info**

No non-base letters.  
Numbers and spaces OK.

Paste Sequence Here

```
1 ttttactctg
cagcagcagc
caccacccat
ggcgctggtt
tcgcgcgcg
tcgtcagtt
61
ctcccggtt
tcacttcctc
acacttcgct
ccactctcac
tctcagcca
cgctctcca
```

Map Sites  
Virtual Digest  
Reset Form

Name your sequence

RestrictionMapper Output - Microsoft Internet Explorer

http://www.restrictionmapper.org/cgi-local/sitefind3.pl

Name: Untitled

Conformation: linear

Overhang: five\_prime, three\_prime, blunt

Minimum Site Length: 6 bases

Maximum Number of Cuts: 2

Included: all commercial, prototypes only

Noncutters: AarI, AatII, AccI, AgeI, AoiI, ApaI, ApaLI, AscI, AsuII, AvrII, BaeI, BamHI, BclI, BglI, BpI, BsePI, BsrBI, BstEII, BtsI, Cfr10I, DraII, DraIII, DrdI, Eam1105I, Eco31I, Eco47III, EcoNI, Esp3I, FseI, FspAI, HaeII, HndII, HpaI, KpnI, MluI, NaeI, NarI, NdeI, NheI, NotI, NruI, OhiI, P1-PspI, PI-SceI, PacI, PmaCI, PmeI, PpiI, PpuMI, PshAI, PstI, PvuI, RsrII, SacII, SalI, SanDI, ScaI, SexAI, SfiI, SgfI, SgrAI, SmaI, SnaBI, SpeI, SphI, SrfI, Sse8387I, SwaI, TaqII, XhoI

Name	Sequence	Site Length	Overhang	Frequency	Cut Positions
<a href="#">BaiI</a>	TGGCCA	6	blunt	1	1512
<a href="#">BsaBI</a>	GATNNNNATC	6	blunt	1	2915
<a href="#">BtrI</a>	CACGTC	6	blunt	1	2560
<a href="#">SbuI</a>	AGGCCT	6	blunt	1	1766
<a href="#">XmnI</a>	GAANNNNTTC	6	blunt	1	1136
<a href="#">AclI</a>	AACGTT	6	five_prime	1	2991
<a href="#">AvaI</a>	CYCGRG	6	five_prime	1	1391
<a href="#">BbvCI</a>	CCTCAGC	7	five_prime	1	1416



## Web Mapping Tools

There are other some free mapping tools on the web  
for finding restriction sites and making text maps,  
but not for nice graphical maps

✳ WebCutter (Max Heiman, Yale Univ.)

<http://www.firstmarket.com/cutter/cut2.html>

✳ EMBOSS Restrict (EMBL Institut Pasteur)

<http://bioweb.pasteur.fr/seqanal/interfaces/restrict.html>

✳ Restriction Maps (Colorado State Univ.)

<http://arbl.cvmbs.colostate.edu/molkit/mapper/index.html>

(uses Java)

## WebCutter

Webcutter is a free on-line tool to restriction map  
nucleotide sequence (text output)

<http://www.firstmarket.com/cutter/cut2.html>

- Webcutter includes the option of finding restriction sites that can be introduced into a sequence by silent mutagenesis

```
NgoAIV          Esp1396I
MroNI           AccB7I
aatggaagccggcgccacctcgctaacggattcaccactccaagaattggagccaatcaattctgcggagaact base pairs
ttacctggccgcccgtggagcgattgcctaagtggtaggttctaacctcggttagttaagaacgcctcttga 1276 to 1350
NgoMI           PflMI
NaeI            Van91I

                AvilI   EcoT14I
Mva1269I Styl Esp1396I
FspI   Eco130I          Gsul
gtgaatgcgcaaccaaccttggcagaacatccatcgctccgcatctccagcagccgcacgcggcgcatc base pairs
cacttacgcggttggtgggaaccgtctgtataggtagcgcaggcggtagaggtcgtcggcgtgcgccgcgtag 1351 to 1425
BsmI   ErhI PflMI          BpmI
BsaMI   BssT1I Van91I
Acc16I   AccB7I
```

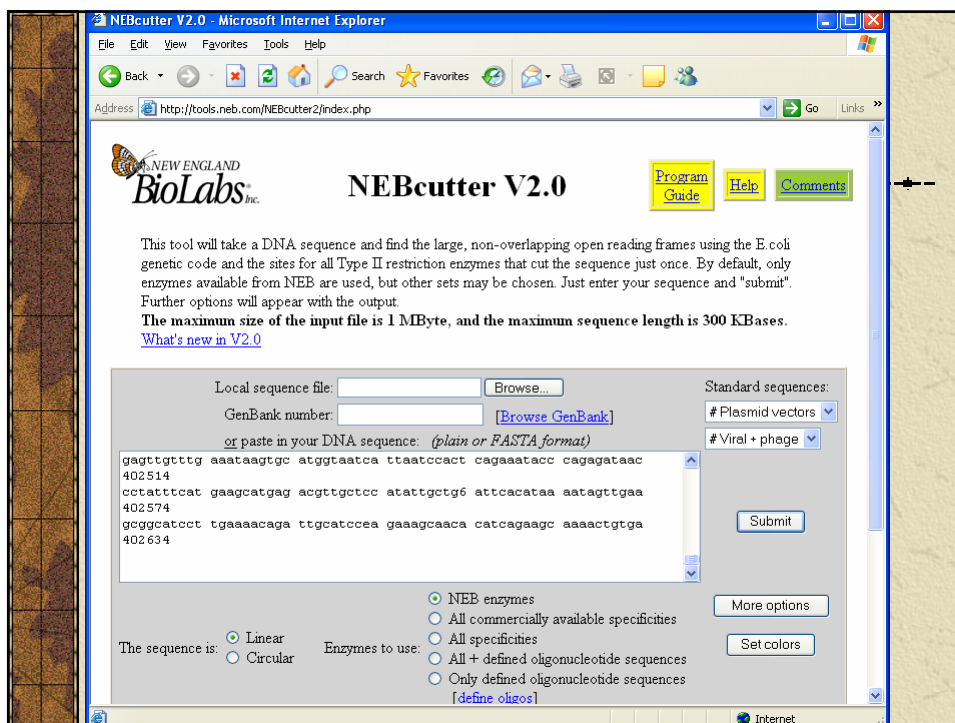
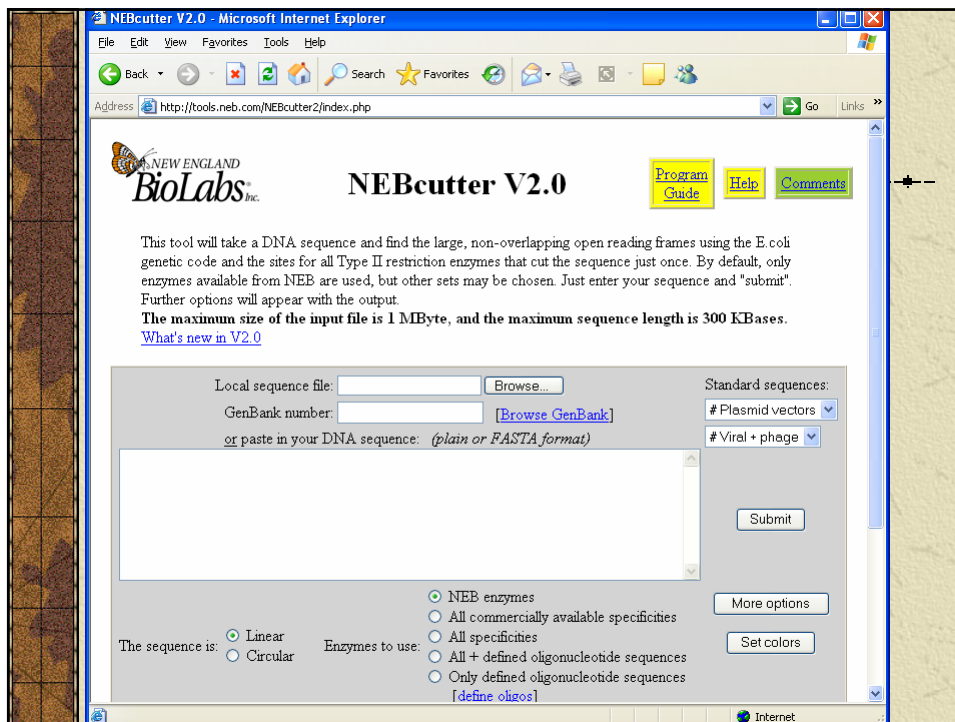
## EMBOSS Restrict

<http://bioweb.pasteur.fr/seqanal/interfaces/restrict.html>

```
# Restrict of , from 1 to 150
# Minimum cuts per enzyme: 1
# Maximum cuts per enzyme: 2000000000
# Minimum length of recognition site: 4
# Blunt ends allowed  # Sticky ends allowed
# DNA is linear      # No ambiguities allowed
# Number of hits: 24
# Base Number  Enzyme      Site      5'  3'  [5'  3']
8  NgoAIV      GCCGGC      8  12
8  NaeI       GCCGGC      10  10
9  MspI       CCGG       9  11
15 AcI       CCGC      12  14
18 MnlI      CCTC      28  27
36 HphI      GGTGA     24  23
45 Tsp509I   AATT      44  48
59 Tsp509I   AATT      58  62
69 AcI       CCGC     66  68
78 BsmI      GAATGC     84  82
81 FspI     TCGCGA     83  83
82 HinfI     GCGC     82  84
82 HhaI     GCGC     84  82
114 Thal     CGCG    115  115
119 AcI       CCGC    119  121
119 HgaI     GACGC    104  109
123 EcoI     GCGCGA    108  106
131 BbvI     GCAGC    143  147
131 BpmI     CTGGAG    111  109
135 AcI       CCGC    135  137
140 Thal     CGCG    141  141
144 AcI       CCGC    141  143
144 HinfI     GCGC    144  146
144 HhaI     GCGC    146  144
```

## NEBcutter

 New England BioLabs







NEBcutter - Microsoft Internet Explorer

Address: <http://tools.neb.com/NEBcutter2/cutshow.php?name=219f4deb->

**Linear Sequence: unnamed sequence**

**Display:** - NEB single cutter restriction enzymes  
- Main non-overlapping, min. 100 aa ORFs  
GC=34%, AT=65%

**Cleavage code**

- blunt end cut
- 5' extension
- 3' extension
- cuts 1 strand

**Enzyme name code**

- Available from NEB
- Has other supplier
- Not commercially available
- \*: cleavage affected by CpG meth.
- :: cleavage affected by other meth.
- (enz.name): ambiguous site

1 12186

Restriction enzymes identified: BpmI, BfrBI, MseI, BsmAI, PpuHI, SacI, BstXI, HfeI, SapI, BstI, NheI, PspOMI, ScaI, BspOI, ClaI, AhdI, PspXI, PaeR7I, TliI, XhoI, BsaBI, BspEI, PvuII, AspCNI, BssSI, BogI.

**Main options**

- New DNA
- Custom digest
- View sequence
- ORF summary

**Availability**

- All commercial
- All

**Display**

- 2 cutters
- 3 cutters

**Zoom**

- Zoom in
- More...

**List**

- 0 cutters
- 1 cutters
- All sites
- Save all sites

NEBcutter - ORF Summary - Microsoft Internet Explorer

Address: <http://tools.neb.com/NEBcutter2/orlist.php?name=219f4deb->

**ORF Summary**

**unnamed sequence**

[\[Back to main display\]](#)

Genetic code: 11. Bacterial and Plant Plastid

Gene	Product	aa seq.	Coordinates	Protein ID	Closest enzyme at 5' end	Closest enzyme at 3' end	Show flanking enzymes
-	-	<a href="#">114 aa</a>	compl(4058..4402)	-	BpmI	BfrBI	<a href="#">show</a>
-	-	<a href="#">107 aa</a>	compl(11569..11892)	-	MseI	BsmAI	<a href="#">show</a>
-	-	<a href="#">98 aa</a>	compl(1657..1953)	-	FokI	MseI	<a href="#">show</a>
-	-	<a href="#">96 aa</a>	9164..9454	-	MscI	AleI	<a href="#">show</a>
-	-	<a href="#">76 aa</a>	compl(10903..11133)	-	DraI	FatI	<a href="#">show</a>
-	-	<a href="#">72 aa</a>	3219..3437	-	MnlI	MseI	<a href="#">show</a>
-	-	<a href="#">70 aa</a>	10107..10319	-	PstI	*ApeKI	<a href="#">show</a>
-	-	<a href="#">63 aa</a>	compl(940..1131)	-	AhlI	HphI	<a href="#">show</a>
-	-	<a href="#">61 aa</a>	compl(2446..2631)	-	BsmFI	MboII	<a href="#">show</a>
-	-	<a href="#">58 aa</a>	compl(6561..6737)	-	SspI	AseI	<a href="#">show</a>
-	-	<a href="#">55 aa</a>	compl(6230..6397)	-	TspRI	Tsp509I	<a href="#">show</a>
-	-	<a href="#">55 aa</a>	7770..7937	-	BsaBI	*BssKI	<a href="#">show</a>
-	-	<a href="#">54 aa</a>	compl(10708..10872)	-	THI	MseI	<a href="#">show</a>
-	-	<a href="#">53 aa</a>	2817..2978	-	BccI	SfaNI	<a href="#">show</a>
-	-	<a href="#">51 aa</a>	compl(2047..2202)	-	HaeIII	BslI	<a href="#">show</a>



NEBcutter - Microsoft Internet Explorer

Address: <http://tools.neb.com/NEBcutter2/cutshow.php?name=219f4deb->

**Linear Sequence: unnamed sequence**

Display: - NEB single cutter restriction enzymes  
- Main non-overlapping, min. 100 aa ORFs  
GC=34%, AT=65%

Cleavage code:  
 ✂ | blunt end cut  
 5' | 5' extension  
 3' | 3' extension  
 | cuts 1 strand

Enzyme name code:  
 Available from NEB  
 Has other supplier  
 Not commercially available  
 \*: cleavage affected by CpG meth.  
 #: cleavage affected by other meth.  
 (enz.name): ambiguous site

1 12186

Restriction enzymes shown: PpuHI, SacI, BstEII, HpaI, \*BseVI, BstXI, \*HfeI, \*FauI, SapI, DrdI, \*BtgZI, BmtI, NheI, BspMI, \*BspEI, PvuII, ApaI, \*AieI, BpuI, PspOMI, \*Scal, \*BspDI, \*ClaI, \*AhdI, \*PspXI, \*PaeR7I, \*TliI, \*XhoI, \*BsaAI, \*BspEI, PvuII, \*AspCNI, BssSI, \*BcgI.

Main options:  
 New DNA  
 Custom digest  
 View sequence  
 ORF summary

Availability:  
 All commercial  
 All

Display:  
 2 cutters  
 3 cutters

Zoom:  
 Zoom in  
 More

List:  
 0 cutters  
 1 cutters  
 All sites

NEBcutter - ORF Sequence - Microsoft Internet Explorer

Address: <http://tools.neb.com/NEBcutter2/orfdisp.php?name=219f4deb-8orfid=0>

**ORF Sequence**

unnamed sequence

[Back to main display]

Coding region: complement(4058..4402)

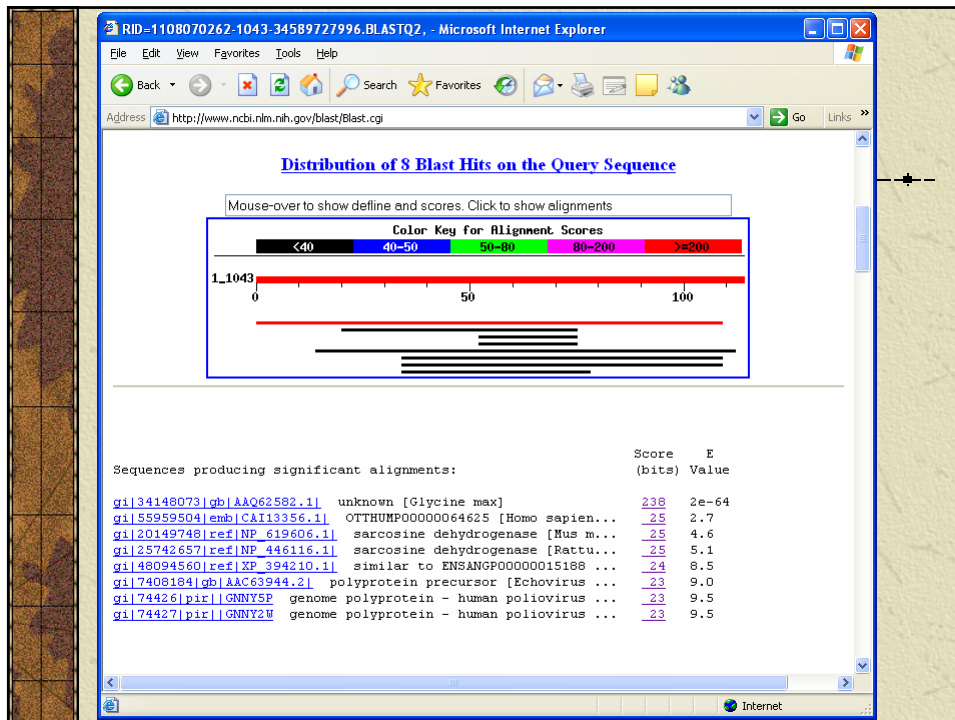
2000 6500

[Edit] - [Delete] - [Add new ORF] - [Locate multiple cutters that excise this ORF] - [Silent Mutagenesis]

Protein sequence:

> 114 aa  
 MEGLMSGPFI FIPYSSVYDH DDAACGTFVS PNEVYVHDST GSIQRMKEFH  
 PQCGSSSPI NKSLCNIYPS LRGFFVDEQ VQEAPPLCSY IQIMLQLSTV  
 TLPSQAADKV SSLE

➔



RID=1108070262-1043-34589727996.BLASTQ2, - Microsoft Internet Explorer

File Edit View Favorites Tools Help

Address <http://www.ncbi.nlm.nih.gov/blast/Blast.cgi> Go Links

### Alignments

>[gi|34148073|gb|AAQ62582.1](#) unknown [Glycine max]  
Length = 2711

Score = 238 bits (607), Expect = 2e-64  
Identities = 110/110 (100%), Positives = 110/110 (100%)

Query: 1 MEGLMSGPFIFIPYSSVYDHDDAACGTFVSPNEVYWHSTGSIQKMEFHPQCGSSSSPI 60  
MEGLMSGPFIFIPYSSVYDHDDAACGTFVSPNEVYWHSTGSIQKMEFHPQCGSSSSPI  
Sbjct: 2044 MEGLMSGPFIFIPYSSVYDHDDAACGTFVSPNEVYWHSTGSIQKMEFHPQCGSSSSPI 2103

Query: 61 NKSLCNIYPSLRGFFVDECQVQEAAPPLCSYIQIMLQLSTVTLPSQAADKV 110  
NKSLCNIYPSLRGFFVDECQVQEAAPPLCSYIQIMLQLSTVTLPSQAADKV  
Sbjct: 2104 NKSLCNIYPSLRGFFVDECQVQEAAPPLCSYIQIMLQLSTVTLPSQAADKV 2153

NEBcutter - Microsoft Internet Explorer

Address: <http://tools.neb.com/NEBcutter2/cutshow.php?name=219f4deb>

**Linear Sequence: unnamed sequence**

Display: - NEB single cutter restriction enzymes  
- Main non-overlapping, min. 100 aa ORFs  
GC=34%, AT=65%

Cleavage code	Enzyme name code
⌵   blunt end cut	Available from NEB
⌵   5' extension	Has other supplier
⌵   3' extension	Not commercially available
⌵   cuts 1 strand	*: cleavage affected by CpG meth.
	⌵: cleavage affected by other meth.
	(enz.name): ambiguous site

1 12186

Restriction enzymes shown: BpmI, BfrBI, MseI, BsmAI, PpuHI, SacI, BstXI, MfeI, SapI, BstI, NheI, HpaI, BstEII, \*FauI, \*BseVI, CspCI, \*BtgZI, DrdI, HpaI, AleI, BpuI, PspOMI, ScaI, \*BspDI, \*C1aI, AhdI, \*PspXI, \*PaeR7I, \*TliI, \*XhoI, BsaBI, \*BspEI, PvuII, \*AspCNI, BssSI, \*BglI.

Main options: New DNA, Custom digest, View sequence, ORF summary

Availability: All commercial, All

Display: 2 cutters, 3 cutters

Zoom: Zoom in, More

List: 0 cutters, 1 cutters, All sites

NEBcutter - ORF Sequence - Microsoft Internet Explorer

Address: <http://tools.neb.com/NEBcutter2/orfdisp.php?name=219f4deb-8&orfid=1>

**ORF Sequence**

unnamed sequence

[Back to main display]

Coding region: complement(11569..11892)

9500 107 aa

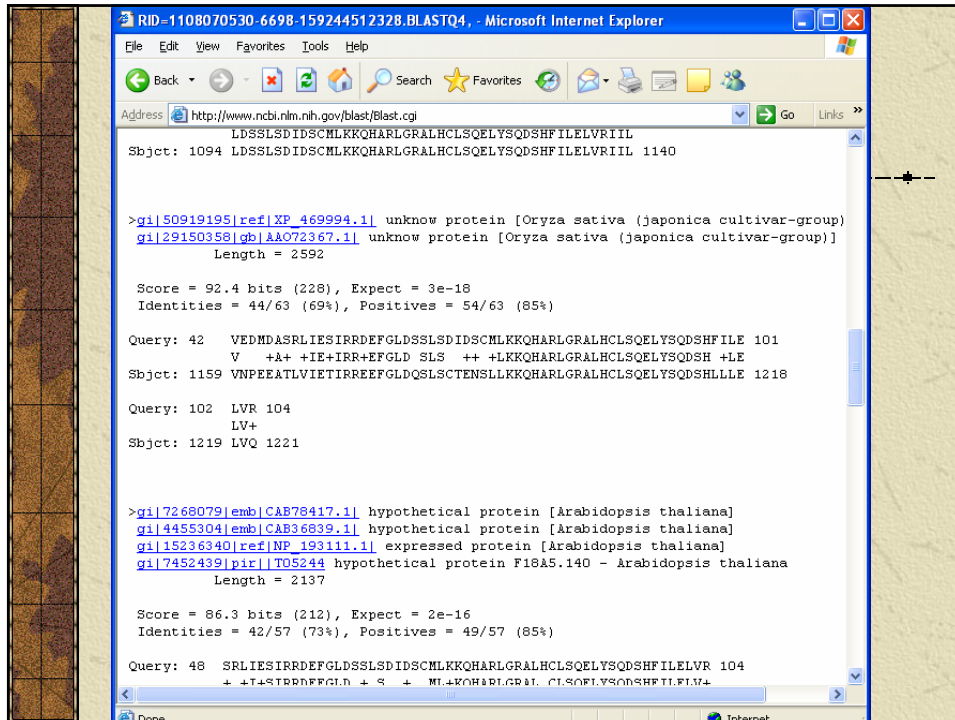
[Edit] - [Delete] - [Add new ORF] - [Locate multiple cutters that excise this ORF] - [Silent Mutagenesis]

Protein sequence:

> 107 aa

MTVSLVPHRL IEGCTEIIET VDPEKSNDES NTCCLGNSFQ HVEDMDASRL  
IESIRRDEFG LDSSLSDIDS CHLKKQHARL GRALHCLSQE LYSQDSHFIL  
ELVRIIL

Blast this sequence at NCBI



## Primer Design

- ✳ Polymerase chain reaction (PCR)
- ✳ DNA sequencing
- ✳ Cloning



## PCR Primer Design

- ✠ The design of PCR (and sequencing) primers is relatively simple from a computational point of view: just search along a sequence and find short sub-sequences that fit certain criteria.
- ✠ However, since the molecular biology of PCR is very complex, the nature of these criteria is not at all obvious.
- ✠ All primers design software uses approximately the same criteria and computing algorithms. Graphical output is not necessary.

## Molecular Biology of PCR

- ✠ The fundamental Molecular Biology of PCR is not well understood.
- ✠ We know what happens in a descriptive sense, but not the physical chemistry/thermodynamics
- ✠ The rules for choosing PCR primers are a rough combination of educated guesses and old fashioned trial-and-error.
- ✠ None of the published formulas for calculating annealing temperatures has been proven to give better than a rough estimate-however, most work!



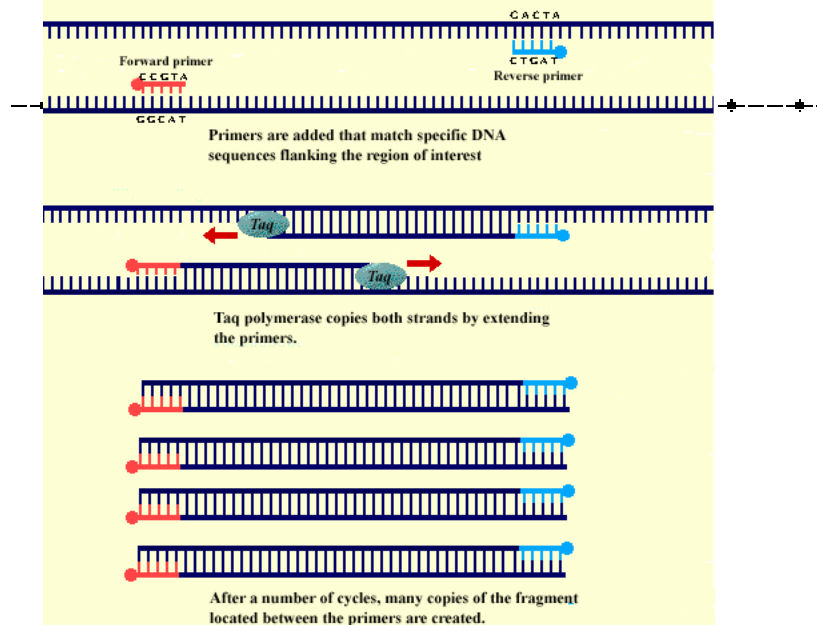
## Polymerase chain reaction

- ✧ Amplify DNA fragment for cloning
- ✧ RT-PCR to measure gene expression
- ✧ Detect polymorphisms for mapping or diagnostics
  
- ✧ Need to design primers from DNA sequence

## How PCR Works

- 
- DNA and two primers are combined in a salt solution with dNTPs and a heat stable DNA polymerase enzyme
  - The primers match some sequence in the target DNA
  - The solution is rapidly heated to DNA denaturing temperatures ( $\sim 95^{\circ}\text{C}$ ) and cooled to a temperature where the polymerase can function
  - Each thermal cycle generates copies of the sequence between the primers, so the total number of fragments amplifies in an exponential fashion: 2, 4, 8, 16, 32, 64, etc.

## Polymerase Chain Reaction



## Primer Design Rules

- ✦ primers should be at least 15 base pairs long
- ✦ have at least 50% G/C content
- ✦ anneal at a temperature in the range of 50-65 degrees C
- ✦ Usually higher annealing temperatures ( $T_m$ ) are better (i.e. more specific for your desired target)
- ✦ forward and reverse primer should anneal at approximately the same temperature

# Primer design sites

## ✧ Primer design sites

- ◆ [http://frodo.wi.mit.edu/cgi-bin/primer3/primer3\\_www.cgi](http://frodo.wi.mit.edu/cgi-bin/primer3/primer3_www.cgi)

## ✧ Primer3

- ◆ [http://frodo.wi.mit.edu/cgi-bin/primer3/primer3\\_www.cgi](http://frodo.wi.mit.edu/cgi-bin/primer3/primer3_www.cgi)

## ✧ GenomeWeb

- ◆ <http://www.hgmp.mrc.ac.uk/GenomeWeb/nuc-primer.html>
- ◆ Web Primer

Primer3 Input (primer3\_www.cgi v 0.2) - Microsoft Internet Explorer

File Edit View Favorites Tools Help

Back Forward Stop Home Search Favorites Media Mail Print

Address [http://frodo.wi.mit.edu/cgi-bin/primer3/primer3\\_www.cgi](http://frodo.wi.mit.edu/cgi-bin/primer3/primer3_www.cgi) Go Links

### Primer3

pick primers from a DNA sequence

Paste source sequence below (5'→3', string of ACGTNacgtm -- other letters treated as N -- numbers and blanks ignored). FASTA format ok. Please N-out undesirable sequence (vector, ALUs, LINES, etc.) or use a [Mispriming Library \(repeat library\)](#): NONE

```
2881 acgtgaattt tgatcgaat gaatgatgc gatggatgc cgaacacatg caaatttgta
2941 ccatctattt atttatcttg attttcttc taggttattt gcttcttata acgtttagt
3001 aatgtaagct tgtttctctt atcttttgc tggaagaata ataagggaat ggaacagaac
3061 tccattaacc ttgattgtga gttacattgt aaaggaacgg aagtaaacag aatttattat
3121 aattttaaac ttgcctaact gtctttttta taaaaaaaaa aaaaaa
```

<input checked="" type="checkbox"/> Pick left primer or use left primer below.	<input type="checkbox"/> Pick hybridization probe (internal oligo) or use oligo below.	<input checked="" type="checkbox"/> Pick right primer or use right primer below (5'→3' on opposite strand).
--	--	---

Pick Primers Reset Form

Sequence Id:  A string to identify your output.

m.  E.g. 50.2 requires primers to surround the 2 bases at positions 50 and 51. Or mark the source sequence with [ and ]:

Internet

Primer3 Input (primer3\_www.cgi v 0.2) - Microsoft Internet Explorer

Address: http://frodo.wi.mit.edu/cgi-bin/primer3/primer3\_www.cgi

### General Primer Picking Conditions

Primer Size: Min: 18 Opt: 20 Max: 27  
 Primer Tm: Min: 57.0 Opt: 60.0 Max: 63.0 Max Tm Difference: 100.0  
 Product Tm: Min: Opt: Max:  
 Primer GC%: Min: 20.0 Opt: Max: 80.0  
 Max Self Complementarity: 8.00 Max 3' Self Complementarity: 3.00  
 Max #N's: 0 Max Poly-X: 5  
 Inside Target Penalty: Outside Target Penalty: 0 Set Inside Target Penalty to allow primers inside a target.  
 First Base Index: 1 CG Clamp: 0  
 Salt Concentration: 50.0 Annealing Oligo Concentration: 50.0 (Not the concentration of oligos in the reaction mix but of those annealing to template.)  
☒ Liberal Base ☐ Show Debugging Info ☒ Do not treat ambiguity codes in libraries as consensus  
 Pick Primers Reset Form

### Other Per-Sequence Inputs

Included Region: E.g. 20,400: only pick primers in the 400 base region starting at position 20. Or use ( and ) in the source sequence to mark the beginning and end of the included region: e.g. in ATC(TTC...TCT)AT the included region is TTC...TCT.  
 Start Codon Position:

### Sequence Quality

## Results

- ✧ Provides best and four other primer pairs
- ✧ Primer sequences
- ✧ Start nt for each primer
- ✧ Length of each primer
- ✧ Tm
- ✧ GC content
- ✧ Size of PCR product



Primer3 Output (primer3\_www\_results.cgi v 0.4) - Microsoft Internet Explorer

Back Forward Stop Search Favorites Media Mail Print

Address [http://frodo.wi.mit.edu/cgi-bin/primer3/primer3\\_www\\_results.cgi](http://frodo.wi.mit.edu/cgi-bin/primer3/primer3_www_results.cgi) Go Links

## Primer3 Output

WARNING: Numbers in input sequence were deleted.

No mispriming library specified  
Using 1-based sequence positions

OLIGO	start	len	tm	gc%	any	3' seq
LEFT PRIMER	1269	20	59.97	50.00	3.00	0.00 gaactggaatggctggtgtt
RIGHT PRIMER	1460	20	60.04	45.00	6.00	2.00 gccattatccaaagcttga

SEQUENCE SIZE: 3166  
INCLUDED REGION SIZE: 3166

PRODUCT SIZE: 192, PAIR ANY COMPL: 5.00, PAIR 3' COMPL: 1.00

```

1 ttttactctgcagcagcagccaccacccatggcgtcggtttccgcgcgcgtcgctcagtt
61 ctcccgcggtttcacctttcacacttcgctccactctcactctcagggcagctcttcca
121 atctcaatgcgcgcctttcttctctctgcgcgaacttccattcactccggaagggtcttac
181 tttaccacggggaagagaggaccgcagtagcatctgtacgtgcttcattacagatgttcc
241 accgaatgtgtccttggaggaaaaacaactacccaaaggagaaacttggtctgttcacaa
301 atttggtggaacctgtgtgggaacctctcagagaataaaaaatgttgccggacataattct

```

## Primer Problems

- ✖ primers should flank the sequence of interest
- ✖ primer sequences should be unique
- ✖ primers that match multiple sequences will give multiple products
- ✖ repeated sequences can be amplified - but only if unique flanking regions can be found where primers can bind
- ✖ primers can have self-annealing regions within each primer (i.e. hairpin and foldback loops)
- ✖ pairs of primers can anneal to each other to form "primer dimers"



## Differential Primers

### ✧ New challenges for PCR primer design

- ◆ gene-specific primers (for multi-gene families)
- ◆ identify specific species or strains of organisms
  - molecular diagnostics/detectors
- ◆ Consensus primers
  - amplify a gene from all of a diverse group of organisms (eg. bacterial 16-S rDNA)

### ✧ Need to work with multiple alignments and find differential or conserved regions

## Primer Design on the Web

### ✧ There are a bunch of good PCR primer design programs on the web:

- ◆ **Primer 3** at the MIT Whitehead Institute  
[http://www.genome.wi.mit.edu/cgi-bin/primer/primer3\\_www.cgi](http://www.genome.wi.mit.edu/cgi-bin/primer/primer3_www.cgi)
- ◆ **Cassandra** at the Univ. of Southern California  
[http://www-hto.usc.edu/software/procrustes/cassandra/cass\\_frm.html](http://www-hto.usc.edu/software/procrustes/cassandra/cass_frm.html)
- ◆ **GeneFisher** by Folker Meyer & Chris Schleiermacher at Bielefeld University, Germany  
<http://bibiserv.TechFak.Uni-Bielefeld.DE/genefisher/>
- ◆ **Xprimer** at the Virtual Genome Center, Univ. Minnesota Medical School  
<http://alces.med.umn.edu/rawprimer.html>

## Other Technologies

### ✠ Multiplex PCR

GeneScan (ABI)

### ✠ PCR related technologies

- ◆ Primer extension
  - Taqman (ABI)
  - Orchid
  - Pyrosequencing
- ◆ Ligase chain reaction
- ◆ Oligos for microarrays
- ◆ DNA sequencing reactions

## RNA secondary structure

- ✠ Look for secondary structures

## What we covered today

- ☐ Restriction site analysis
  - ☐ Cloning
  - ☐ Southern blot analysis
- ☐ Primer design
  - ☐ Cloning, amplification gene expression, sequencing